

Review

Open Access

Highlights on endoglin (CD105): from basic findings towards clinical applications in human cancer

Ester Fonsatti¹ and Michele Maio^{*1,2}

Address: ¹Cancer Bioimmunotherapy Unit, Department of Medical Oncology, Centro di Riferimento Oncologico, I.R.C.C.S., 33081 Aviano, Italy and ²Division of Medical Oncology and Immunotherapy, Department of Oncology, University Hospital of Siena, 53100 Siena, Italy

Email: Ester Fonsatti - efonsatti@cro.it; Michele Maio* - mmaio@cro.it

* Corresponding author

Published: 11 June 2004

Received: 07 April 2004

Accepted: 11 June 2004

Journal of Translational Medicine 2004, **2**:18

This article is available from: <http://www.translational-medicine.com/content/2/1/18>

© 2004 Fonsatti and Maio; licensee BioMed Central Ltd. This is an Open Access article: verbatim copying and redistribution of this article are permitted in all media for any purpose, provided this notice is preserved along with the article's original URL.

Abstract

Antibody targeting of tumor-associated vasculature is a promising therapeutic approach in human cancer; however, a specific cell membrane marker for endothelial cells of tumor vasculature has not been discovered yet. Endoglin (CD105) is a cell-surface glycoprotein most recently identified as an optimal indicator of proliferation of human endothelial cells. The finding that CD105 is over-expressed on vascular endothelium in angiogenetic tissues has prompted several pre-clinical studies designed to get a deeper understanding on the role of CD105 in angiogenesis, and to evaluate the most appropriate clinical setting(s) to utilize CD105 as a therapeutic target. In this review, the foreseeable clinical applications of CD105 in human cancer are discussed.

Background

The availability of new and more sophisticated technologies, together with the improved knowledge on tumor-host interactions, have allowed the identification and characterization of different tumor-associated antigens (TAA) to be used as molecular targets for immunotherapeutic approaches in patients with solid or hematologic malignancies. Prompted by encouraging pre-clinical evidences, significant clinical results in cancer treatment have been obtained through antibody-based therapeutic regimens, such as those that target CD20 on malignant B cells [1] or HER2 in breast cancer [2]. However, due to the heterogeneous expression of TAA in neoplastic tissues, these approaches raise some critical issues such as "patients' eligibility" to specific TAA-based treatment modalities. Moreover, the efficacy of TAA targeting is frequently limited by the inadequate accessibility of therapeutic antibodies or their derived molecules within the tumor mass [3].

Currently, great interest is focused on angiogenesis and on its potential clinical implications in cancer, and vascular targeting represents a highly promising alternative to the direct engagement of therapeutic TAA on neoplastic cells [4,5]. Among potential therapeutic strategies to induce tumor regression by blocking tumor blood supply, an intriguing approach relies on the selective targeting of cell surface molecules over-expressed on endothelial cells of tumor-associated blood vessels [4,5]. In this setting, emerging *in vitro* and *in vivo* pre-clinical evidence identifies CD105 as a cell membrane glycoprotein representing a prime vascular target to implement innovative antibody-based diagnostic and therapeutic strategies shared by human neoplasia of different histotype.

Biological features of CD105

Tissue distribution

CD105 is a 180 kDa transmembrane glycoprotein constitutively phosphorylated [6-10], with a marked tissue-specificity [11]. Supporting this notion, CD105 is

Table I: In vivo distribution of CD105 on non-endothelial cells.

Histotype
Activated monocytes
Differentiated macrophages
Early B cells
Erythroid precursors
Fibroblasts
Follicular dendritic cells
Melanocytes
Heart mesenchimal cells
Vascular smooth muscle cells
Mesangial cells
Syncytiotrophoblasts

predominantly expressed on endothelial cells [11-13] and its promoter is strongly and selectively active in endothelial cells [14,15]. Consistently, elevated levels of CD105 expression were detected on human microvascular endothelium [16] and on vascular endothelial cells in tissues undergoing active angiogenesis, such as regenerating and inflamed tissues or tumors [11,12,17-21]. However, CD105 was also weakly expressed on selected non-endothelial cells of different histotype (Table 1 and ref [22,23] for review).

In solid neoplasia, CD105 is present on endothelial cells of both peri- and intra-tumoral blood vessels and on tumor stromal components [11,17,22-24]. In particular, CD105 is largely expressed in small and likely immature tumor vessels as demonstrated in breast, prostate and gastric cancer [24-26]; rarely, CD105 is expressed in the cytoplasm of neoplastic cells [23]. In lung carcinoma, staining for CD105 was reported to be strong at the areas of active angiogenesis including tumor edge, while it was less intense in the central area of the tumor and not detectable in the adjacent normal tissue [12].

Functional activity

CD105 is a component of the receptor complex of Transforming Growth Factor (TGF)- β [27-29], a pleiotropic cytokine involved in cellular proliferation, differentiation and migration [30]. It binds several components of the TGF- β superfamily [27,29]. Interestingly, binding of TGF- β 1 to CD105 reduces the levels of CD105 phosphorylation [10] and the levels of CD105 expression modulate the effects of TGF- β 1 [28,31-35]. In this respect, it is of interest that the inhibition of CD105 expression enhanced the ability of TGF- β 1 to suppress growth, migration and capacity to form capillary tubes of cultured endothelial cells [32].

In the absence of TGF- β 1, CD105 shows an anti-apoptotic effect in endothelial cells under hypoxic stress, suggesting

for a protective role of CD105 against pro-apoptotic factors [36].

In addition, the discovery that levels of CD105 regulate the expression of different components of the extracellular matrix including fibronectin, collagen, PAI-1 and lumican [34,37,38], is also suggestive for a crucial role of CD105 in cellular transmigration [38].

Modulation

Different environmental factors and cytokines involved in angiogenesis modulate CD105 expression. The levels of CD105 protein, mRNA and promoter activity are up-regulated by hypoxia [39] and by TGF- β 1 [28,39-41], which cooperate to induce the expression of CD105 at transcriptional level [39]. Instead, TNF- α down-regulates CD105 protein levels but it has no effect at the transcriptional level [42].

Furthermore, CD105 expression was up-regulated on human umbilical vein endothelial cells (HUVEC) infected with a recombinant adenovirus carrying a constitutively active form of activin receptor-like kinase (ALK)-1, a type I TGF- β receptor [43].

Supporting the *in vivo* modulation of CD105 by pro-angiogenic stimuli, elevated levels of CD105 were associated with high levels of vascular endothelial growth factor in non-small cell lung cancer lesions positive for angiopoietin-2, a regulatory factor of survival of endothelial cells, considerably expressed at sites of vascular remodelling and in highly vascularized tumors [44].

CD105 and vascularization

Even if its functional role is not fully understood, several findings suggest for the involvement of CD105 in angiogenesis and vascular development, and in maintaining vessel wall integrity.

Table 2: Intra-tumor microvessel density determined by immunohistochemical staining for CD105: an indicator of poor prognosis in patients with solid neoplasia.

Tumor histotype	References
Breast carcinoma	[54-55, 75-76]
Cervical cancer	[77]
Colorectal cancer	[78-79]
Endometrial carcinoma	[80-81]
Gastric carcinoma	[26]
Melanoma	[82]
Nonseminomatous testicular germ cell tumors	[83]
Non-small cell lung cancer	[56, 84]
Prostate cancer	[25]
Renal cell carcinoma	[85]
Squamous cell carcinoma of the oral cavity	[19]

Table 3: CD105 as a marker of survival in patients with solid tumors of different histotype.

Tumor histotype	CD105-MVD ^a correlation with survival	References
Breast carcinoma	The number of CD105-positive microvessels correlated significantly ($p = 0.001$) with poor overall survival.	[55]
Colorectal cancer	Patients with CD105-MVD above the median showed the worst prognosis; similar results were obtained when CD105-MVD was divided in quartiles.	[78]
Endometrial carcinoma	Patients with the lower quartiles for CD105-MVD showed reduced survival compared to those with the higher quartiles.	[80]
Non-small cell lung cancer	5-year survival rate of patients with the lower CD105-MVD was higher compared to that of patients with the higher CD105-MVD.	[84]
Prostate cancer	Median survival time were shorter for patients with CD105-MVD above the median.	[25]

^aCD105-MVD, intratumor microvascular density as determined by anti-CD105 monoclonal antibodies.

First, CD105 expression is up-regulated on proliferating endothelial cells in culture [11-13] and on endothelial cells of angiogenetic blood vessels [11,12,24,33,45]. Furthermore, CD105 knockout mice died of defective vascular development during early gestation [46], as observed in *TGF-β1*- and *ALK-1*-null mice [47,48]. In particular, CD105 null mice showed important structural defects in the primitive vascular plexus of the yolk sac that prevented the formation of normal mature vessels [46]. Additionally, both in humans and mice, CD105 gene mutations are associated with hereditary hemorrhagic telangiectasia type 1, an inherited disease characterized by arteriovenous malformations and bleedings [49-51]. Finally, CD105 has been most recently suggested as a regulator factor of nitric oxide-dependent vasodilatation. In fact, the levels of CD105 expression modulated the amounts of endothelial nitric oxide synthase (eNOS) in kidney and femoral arteries of mice. Furthermore, over-expression or suppression of CD105 in cultured endothelial cells induced a marked increase or decrease in the protein levels of eNOS, respectively [52].

Interestingly, an increment in microvessel density, as determined by immunohistochemical staining for CD105, was found during the progressive stages of colorectal carcinogenesis [53]. In line with this finding, the assessment of neovascularization by CD105 staining was found to represent a potential predictor of prognosis in different solid malignancies (Table 2 and Table 3). For instance, the CD105-positive blood vessels count was prognostic for survival in patients with prostate cancer of Gleason score 5-7 [25], and correlated with overall survival of node-negative patients affected by breast carcinoma [54,55].

mAb directed to CD105 but not to the pan-endothelial marker CD34 revealed an inverse correlation between intra-tumoral microvessel density and apoptotic index of neoplastic cells in non-small cell lung cancer patients [56]. Since angiogenesis is crucial for tumor development and progression [57], this finding provided supportive evidence to the usefulness of CD105 targeting in antiangiogenic therapy of cancer [56].

CD105 targeting

Ex vivo background

Selected anti-CD105 monoclonal antibodies (mAb) significantly inhibit the proliferation of cultured human microvascular and macrovascular endothelial cells [11,58,59], thus supporting the notion that CD105 is a promising vascular target to implement innovative antibody-based therapeutic strategies in human cancer. Noteworthy, differences in the growth suppression of endothelial cells have been found among 4 anti-CD105 mAb defining different epitopes [59]; nevertheless, TGF- β 1 and each of the 4 anti-CD105 mAb showed synergistic suppression of HUVEC proliferation [59].

A bispecific single-chain diabody directed to the adenovirus fiber knob domain and to CD105 was shown to be effective in enhancing adenovirus transduction in HUVEC, thus sustaining the use of CD105 protein as target for therapeutic gene transfer in endothelial cells [60]. Along this line, a vector constructed with the CD105 promoter was efficiently utilized to deliver gene expression specifically to endothelial cells of mouse blood vessels [61]. Additionally, human CD105 promoter fragments were successfully utilized in pigs to drive CD59 expression in the small vessels of heart, kidney and lung, but not in the large vessels of these organs [62].

Most recently, another bispecific single-chain diabody was proposed for therapeutic approaches aiming to destroy tumor-associated vasculature. This engineered antibody is directed to human CD105 as well as CD3 and it is effective to mediate killing of CD105-positive endothelial cells by cytotoxic T lymphocytes [63]. An alternative strategy of CD105 targeting for anti-angiogenic treatment of cancer might derive by the use of conditionally replicating adenoviruses (CRAD). In fact, CRAD obtained by utilizing Flk-1 and CD105 regulatory elements have been transcriptionally targeted towards proliferating endothelial cells, with specificity and efficacy in killing HUVEC [64].

In vivo diagnostic targeting

The over-expression of CD105 on proliferating endothelial cells of the tumour vasculature suggested that CD105 might also represent a good target for the immunoscintigraphy of tumors. In keeping with this idea, targeting of CD105 by radiolabeled mAb was described as a safe and effective procedure to image tumors in animal models [13,65]. The intravenous administration of a ^{125}I -labeled anti-CD105 mAb efficiently imaged spontaneous mammary adenocarcinomas in dogs. The immunoscintigraphy performed 8 hours after mAb injection demonstrated that the uptake of the radiolabeled mAb was rapid and intense, and no systemic side effects were observed in the injected dogs during a 3 months follow-up after imaging proce-

dures [13]. Consistently, the scintigraphy performed 15 minutes after administration of low doses of ^{111}In -labeled anti-CD105 mAb in C57BL/6 mice demonstrated an accumulation of radioactivity in xenografts of human melanoma. The autoradiography and immunohistology showed a marked concentration of the mAb in the periphery of the tumor mass with an heterogeneous distribution in its centre. Noteworthy, the 97% of the injected dose of the radiolabeled anti-CD105 mAb was removed from the circulation within 15 min, and the blood half-life of the anti-CD105 mAb was estimated to be <1 minute [65].

The immunoscintigraphy performed after renal artery perfusion of ^{99}Tcm -labeled anti-CD105 mAb in the freshly excised kidney from a patient with renal carcinoma identified 2 distinct hot spots of radioactivity, which matched the positions of the tumors, as demonstrated by the subsequent histopathologic examination; noteworthy, only one of the two tumor masses was identified by a pre-surgery magnetic resonance imaging scan [66].

In vivo therapeutic targeting

Targeting of CD105, as therapeutic antiangiogenic approach in cancer, has been extensively investigated in severe combined immunodeficiency [SCID] mice bearing human breast tumors. The results of these studies demonstrated a long lasting suppression of tumor growth and metastasis by systemic administration of radiolabeled or immunotoxin-conjugated anti-CD105 mAb [67-69]. Furthermore, naked anti-CD105 mAb, which reacted strongly with proliferating human endothelial cells but weakly with murine endothelial cells, showed synergism with conventional chemotherapeutic regimens in a human skin/SCID mouse chimera model [70]. Interestingly, in all these animal models the anti-tumor efficacy and the anti-metastatic activities were identified in the ability of the anti-CD105 mAb to inhibit tumor-associated angiogenesis and/or to obliterate tumor-associated vasculature [67-70].

Conclusions and future directions

A number of convincing experimental findings suggest that selected anti-CD105 mAb can strongly localize to the endothelium of tumor-associated vasculature and that they are efficient to inhibit tumor angiogenesis, tumor growth and metastasis in mice, pointing to CD105 as a suitable vascular target to implement antibody-based therapeutic approaches in cancer.

However, concern about the therapeutic applications of anti-CD105 mAb and their derived molecules in cancer patients emerged by discrepancies observed in the expression of CD105 within normal and tumor tissues [71-74]. In this respect, it has been suggested that not all anti-CD105 mAb are useful for anti-angiogenic targeting

since different mAb have different reactivity with the vasculature of normal tissues [45,59,66,73]. Based on these findings, the comparative evaluation of the reactivity of a large panel of anti-CD105 mAb in the same tumor specimen has been proposed to identify the most reactive with tumor endothelium [45].

Nevertheless, the information on CD105 so far obtained by *ex vivo* studies and in animal models warrants additional efforts to further define the most appropriate therapeutic setting [s] for CD105 in human cancer, and to translate pre-clinical evidences into phase I/II clinical trials.

Abbreviations

ALK, activin receptor-like kinase

CRAD, conditionally replicating adenoviruses

eNOS, endothelial nitric oxide synthase

HUVEC, human umbilical vein endothelial cells

TAAs, tumor-associated antigens

TGF, transforming growth factor

Acknowledgments

This work was supported in part by the Associazione Italiana per la Ricerca sul Cancro and by the progetto Ricerca Finalizzata awarded by the Italian Ministry of Public Health.

References

- Smith MR: **Rituximab (monoclonal anti-CD20 antibody): mechanisms of action and resistance.** *Oncogene* 2003, **22**:7359-7368.
- Ménard S, Pupa SM, Campiglio M, Tagliabue E: **Biologic and therapeutic role of HER2 in cancer.** *Oncogene* 2003, **22**:6570-6578.
- Green MC, Murray JL, Hortobagyi GN: **Monoclonal antibody therapy for solid tumors.** *Cancer Treat Rev* 2000, **26**:269-286.
- Brekken RA, Li C, Kumar S: **Strategies for vascular targeting in tumors.** *Int J Cancer* 2002, **100**:123-130.
- Thorpe PE: **Vascular targeting agents as cancer therapeutics.** *Clin Cancer Res* 2004, **10**:415-427.
- Haruta Y, Seon BK: **Distinct human leukemia-associated cell surface glycoprotein GP160 defined by monoclonal antibody SN6.** *Proc Natl Acad Sci U S A* 1986, **83**:7898-7902.
- Gougos A, Letarte M: **Identification of a human endothelial cell antigen with monoclonal antibody 44G4 produced against a pre-B leukemic cell line.** *J Immunol* 1988, **141**:1925-1933.
- Gougos A, Letarte M: **Primary structure of endoglin, an RGD-containing glycoprotein of human endothelial cells.** *J Biol Chem* 1990, **265**:8361-8364.
- Bellón T, Corbi A, Lastres P, Calés C, Cebrán M, Vera S, Cheifetz S, Massagué J, Letarte M, Bernabéu C: **Identification and expression of two forms of the human transforming growth factor-beta-binding protein endoglin with distinct cytoplasmic regions.** *Eur J Immunol* 1993, **23**:2340-2345.
- Lastres P, Martín-Perez J, Langa C, Bernabéu C: **Phosphorylation of the human-transforming-growth-factor-β-binding protein endoglin.** *Biochem J* 1994, **301**:765-768.
- Burrows FJ, Derbyshire EJ, Tazzari PL, Amlot P, Gazdar AF, King SW, Letarte M, Vitetta ES, Thorpe PE: **Up-regulation of endoglin on vascular endothelial cells in human solid tumors: implications for diagnosis and therapy.** *Clin Cancer Res* 1995, **1**:1623-1634.
- Miller DW, Graulich W, Karges B, Stahl S, Ernst M, Ramaswamy A, Sedlacek HH, Müller R, Adamkiewicz J: **Elevated expression of endoglin, a component of the TGF-β-receptor complex, correlates with proliferation of tumor endothelial cells.** *Int J Cancer* 1999, **81**:568-572.
- Fonsatti E, Jekunen AP, Kairemo KJ, Coral S, Snellman M, Nicotra MR, Natali PG, Altomonte M, Maio M: **Endoglin is a suitable target for efficient imaging of solid tumors: in vivo evidence in a canine mammary carcinoma model.** *Clin Cancer Res* 2000, **6**:2037-2043.
- Graulich W, Nettelbeck DM, Fischer D, Kissel T, Muller R: **Cell type specificity of the human endoglin promoter.** *Gene* 1999, **227**:55-62.
- Botella LM, Sanchez-Elsner T, Sanz-Rodríguez F, Kojima S, Shimada J, Guerrero-Esteo M, Cooreman MP, Ratziu V, Langa C, Vary CP, Ramirez JR, Friedman S, Bernabeu C: **Transcriptional activation of endoglin and transforming growth factor-beta signaling components by cooperative interaction between Sp1 and KLF6: their potential role in the response to vascular injury.** *Blood* 2002, **100**:4001-4010.
- Wong SH, Hamel L, Chevalier S, Philip A: **Endoglin expression on human microvascular endothelial cells association with betaglycan and formation of higher order complexes with TGF-beta signalling receptors.** *Eur J Biochem* 2000, **267**:5550-5560.
- Wang JM, Kumar S, Pye D, van Agthoven AJ, Krupinski J, Hunter RD: **A monoclonal antibody detects heterogeneity in vascular endothelium of tumours and normal tissues.** *Int J Cancer* 1993, **54**:363-370.
- Krupinski J, Kaluza J, Kumar P, Kumar S, Wang JM: **Role of angiogenesis in patients with cerebral ischemic stroke.** *Stroke* 1994, **25**:1794-1798.
- Schimming R, Marre D: **Endoglin (CD105) expression in squamous cell carcinoma of the oral cavity.** *Head Neck* 2002, **24**:151-156.
- Torsney E, Charlton R, Parums D, Collis M, Arthur HM: **Inducible expression of human endoglin during inflammation and wound healing in vivo.** *Inflamm Res* 2002, **51**:464-470.
- Chung YC, Hou YC, Pan AC: **Endoglin (CD105) expression in the development of haemorrhoids.** *Eur J Clin Invest* 2004, **34**:107-112.
- Fonsatti E, Del Vecchio L, Altomonte M, Sigalotti L, Nicotra MR, Coral S, Natali PG, Maio M: **Endoglin: An accessory component of the TGF-beta-binding receptor-complex with diagnostic, prognostic, and bioimmunotherapeutic potential in human malignancies.** *J Cell Physiol* 2001, **188**:1-7.
- Fonsatti E, Altomonte M, Nicotra MR, Natali PG, Maio M: **Endoglin (CD105): a powerful therapeutic target on tumor-associated angiogenetic blood vessels.** *Oncogene* 2003, **22**:6557-6563.
- Wang JM, Kumar S, Pye D, Haboubi N, Al-Nakib L: **Breast carcinoma: comparative study of tumor vasculature using two endothelial cell markers.** *J Natl Cancer Inst* 1994, **86**:386-388.
- Wikstrom P, Lissbrant IF, Stattin P, Egevad L, Bergh A: **Endoglin [CD105] is expressed on immature blood vessels and is a marker for survival in prostate cancer.** *Prostate* 2002, **51**:268-275.
- Yu JX, Zhang XT, Liao YQ, Zhang QY, Chen H, Lin M, Kumar S: **Relationship between expression of CD105 and growth factors in malignant tumors of gastrointestinal tract and its significance.** *World J Gastroenterol* 2003, **9**:2866-2869.
- Cheifetz S, Bellón T, Calés C, Vera S, Bernabéu C, Massagué J, Letarte M: **Endoglin is a component of the transforming growth factor-β receptor system in human endothelial cells.** *J Biol Chem* 1992, **267**:19027-19030.
- Lastres P, Letamendia A, Zhang H, Ríus C, Almendro N, Raab U, López LA, Langa C, Fabra A, Letarte M, Bernabéu C: **Endoglin modulates cellular response to TGF-β1.** *J Cell Biol* 1996, **133**:1109-1121.
- Barbara NP, Wray JL, Letarte M: **Endoglin is an accessory protein that interacts with the signaling receptor complex of multiple members of the transforming growth factor-β superfamily.** *J Biol Chem* 1999, **274**:584-594.
- Govindan R, Bhoola KD: **Genealogy, expression, and cellular function of transforming growth factor-beta.** *Pharmacol Ther* 2003, **98**:257-265.

31. Letamendia A, Lastres P, Botella LM, Raab U, Langa C, Velasco B, Attisano L, Bernabeu C: **Role of endoglin in cellular responses to transforming growth factor- β .** *J Biol Chem* 1998, **273**:33011-33019.
32. Li C, Hampson IN, Hampson L, Kumar P, Bernabeu C, Kumar S: **CD105 antagonizes the inhibitory signaling of transforming growth factor- β 1 on human vascular endothelial cells.** *FASEB J* 2000, **14**:55-64.
33. Ma X, Labinaz M, Goldstein J, Millere H, Keon WJ, Letarte M, O'Brien E: **Endoglin is overexpressed after arterial injury and is required for transforming growth factor- β -induced inhibition of smooth muscle cell migration.** *Arterioscler Thromb Vasc Biol* 2000, **20**:2546-2552.
34. Diez-Marques L, Ortega-Velazquez R, Langa C, Rodriguez-Barbero A, Lopez-Novoa JM, Lamas S, Bernabeu C: **Expression of endoglin in human mesangial cells: modulation of extracellular matrix synthesis.** *Biochim Biophys Acta* 2002, **1587**:36-44.
35. Guerrero-Esteo M, Sanchez-Elsner T, Letamendia A, Bernabeu C: **Extracellular and cytoplasmic domains of endoglin interact with the transforming growth factor-beta receptors I and II.** *J Biol Chem* 2002, **277**:29197-29209.
36. Li C, Issa R, Kumar P, Hampson IN, Lopez-Novoa JM, Bernabeu C, Kumar S: **CD105 prevents apoptosis in hypoxic endothelial cells.** *J Cell Sci* 2003, **116**:2677-2685.
37. Guerrero-Esteo M, Lastres P, Letamendia A, Perez-Alvarez MJ, Langa C, Lopez LA, Fabra A, Garcia-Pardo A, Vera S, Letarte M, Bernabeu C: **Endoglin overexpression modulates cellular morphology, migration, and adhesion of mouse fibroblasts.** *Eur J Cell Biol* 1999, **78**:614-623.
38. Botella LM, Sanz-Rodriguez F, Sanchez-Elsner T, Langa C, Ramirez JR, Vary C, Roughley PJ, Bernabeu C: **Lumican is down-regulated in cells expressing endoglin. Evidence for an inverse correlation between Endoglin and Lumican expression.** *Matrix Biol* 2004, **22**:561-572.
39. Sanchez-Elsner T, Botella LM, Velasco B, Langa C, Bernabeu C: **Endoglin expression is regulated by transcriptional cooperation between the hypoxia and transforming growth factor-beta pathways.** *J Biol Chem* 2002, **277**:43799-43808.
40. Rius C, Smith JD, Almendro N, Langa C, Botella LM, Marchuk DA, Vary CPH, Bernabeu C: **Cloning of the promoter region of human endoglin, the target gene for hereditary hemorrhagic telangiectasia type I.** *Blood* 1998, **92**:4677-4690.
41. Zhu Y, Sun Y, Xie L, Jin K, Sheibani N, Greenberg DA: **Hypoxic induction of endoglin via mitogen-activated protein kinases in mouse brain microvascular endothelial cells.** *Stroke* 2003, **34**:2483-8.
42. Li C, Guo B, Ding S, Rius C, Langa C, Kumar P, Bernabeu C, Kumar S: **TNF alpha down-regulates CD105 expression in vascular endothelial cells: a comparative study with TGF beta I.** *Anticancer Res* 2003, **23**:1189-1196.
43. Ota TM, Fujii M, Sugizaki T, Ishii M, Miyazawa K, Aburatani H, Miyazono K: **Targets of transcriptional regulation by two distinct type I receptors for transforming growth factor-beta in human umbilical vein endothelial cells.** *J Cell Physiol* 2002, **193**:299-318.
44. Tanaka F, Ishikawa S, Yanagihara K, Miyahara R, Kawano Y, Li M, Otake Y, Wada H: **Expression of angiopoietins and its clinical significance in non-small cell lung cancer.** *Cancer Res* 2002, **62**:7124-7129.
45. Duff SE, Li C, Garland JM, Kumar S: **CD105 is important for angiogenesis: evidence and potential applications.** *FASEB J* 2003, **17**:984-992.
46. Li DY, Sorensen LK, Brooke BS, Urness LD, Davis EC, Taylor DG, Boak BB, Wendel DP: **Defective angiogenesis in mice lacking endoglin.** *Science* 1999, **284**:1534-1537.
47. Dickson MC, Martin JS, Cousins FM, Kulkarni AB, Karlsson S, Akhurst RJ: **Defective haematopoiesis and vasculogenesis in transforming growth factor-beta I knock out mice.** *Development* 1995, **121**:1845-54.
48. Urness LD, Sorensen LK, Li DY: **Arteriovenous malformations in mice lacking activin receptor-like kinase-I.** *Nat Genet* 2000, **26**:328-331.
49. McAllister KA, Grogg KM, Johnson DW, Gallione CJ, Baldwin MA, Jackson CE, Helmbold EA, Markef DS, McKinnon WC, Murrell J, McCormick MK, Pericak-Vance MA, Heutink P, Ooststra BA, Haitjema T, Westerman CJ, Porteous ME, Guttmacher AE, Letarte M, Marck DA: **Endoglin, a TGF-beta binding protein of endothelial cells, is the gene for hereditary haemorrhagic telangiectasia type I.** *Nat Genet* 1994, **8**:345-351.
50. Bourdeau A, Faughnan ME, McDonald ML, Paterson AD, Wanless IR, Letarte M: **Potential role of modifier genes influencing transforming growth factor-beta1 levels in the development of vascular defects in endoglin heterozygous mice with hereditary hemorrhagic telangiectasia.** *Am J Pathol* 2001, **158**:2011-2020.
51. van den Driesche S, Mummary CL, Westermann CJ: **Heredity hemorrhagic telangiectasia: an update on transforming growth factor beta signaling in vasculogenesis and angiogenesis.** *Cardiovasc Res* 2003, **58**:20-31.
52. Jerkic M, Rivas-Elena JV, Prieto M, Carron R, Sanz-Rodriguez F, Perez-Barriocanal F, Rodriguez-Barbero A, Bernabeu C, Lopez-Novoa JM: **Endoglin regulates nitric oxide-dependent vasodilatation.** *FASEB J* 2004, **18**:609-611.
53. Akagi K, Ikeda Y, Sumiyoshi Y, Kimura Y, Kinoshita J, Miyazaki M, Abe T: **Estimation of angiogenesis with anti-CD105 immunostaining in the process of colorectal cancer development.** *Surgery* 2002, **131**(1 Suppl):S109-113.
54. Dales JP, Garcia S, Carpenter S, Andrac L, Ramuz O, Lavaut MN, Allasia C, Bonnier P, Taranger-Charpin C: **Long-term prognostic significance of neoangiogenesis in breast carcinomas: comparison of Tie-2/Tek, CD105, and CD31 immunocytochemical expression.** *Hum Pathol* 2004, **35**:176-183.
55. Dales JP, Garcia S, Carpenter S, Andrac L, Ramuz O, Lavaut MN, Allasia C, Bonnier P, Taranger-Charpin C: **Prediction of metastasis risk [11 year follow-up] using VEGF-R1, VEGF-R2, Tie-2/Tek and CD105 expression in breast cancer [n = 905].** *Br J Cancer* 2004, **90**:1216-1221.
56. Tanaka F, Otake Y, Yanagihara K, Kawano Y, Miyahara R, Li M, Ishikawa S, Wada H: **Correlation between apoptotic index and angiogenesis in non-small cell lung cancer: comparison between CD105 and CD34 as a marker of angiogenesis.** *Lung Cancer* 2003, **39**:289-296.
57. Carmeliet P, Jain RK: **Angiogenesis in cancer and other diseases.** *Nature* 2000, **407**:249-257.
58. Maier JA, Delia D, Thorpe PE, Gasparini G: **In vitro inhibition of endothelial cell growth by the antiangiogenic drug AGM-1470 [TNP-470] and the anti-endoglin antibody TEC-11.** *Anticancer Drugs* 1997, **8**:238-244.
59. She X, Matsuno F, Harada N, Tsai H, Seon BK: **Synergy between anti-endoglin [CD105] monoclonal antibodies and TGF-beta in suppression of growth of human endothelial cells.** *Int J Cancer* 2004, **108**:251-257.
60. Nettelbeck DM, Miller DW, Jerome V, Zuzarte M, Watkins SJ, Hawkins RE, Muller R, Kontermann RE: **Targeting of adenovirus to endothelial cells by a bispecific single-chain diabody directed against the adenovirus fiber knob domain and human endoglin [CD105].** *Mol Ther* 2001, **3**:882-891.
61. Velasco B, Ramirez JR, Rellosa M, Li C, Kumar S, Lopez-Bote JP, Perez-Barriocanal F, Lopez-Novoa JM, Cowan PJ, d'Apice AJ, Bernabeu C: **Vascular gene transfer driven by endoglin and ICAM-2 endothelial-specific promoters.** *Gene Ther* 2001, **8**:897-904.
62. Cowan PJ, Shinkel TA, Fiscardo N, Godwin JW, Bernabeu C, Almendro N, Rius C, Lonie AJ, Nottle MB, Wigley PL, Paizis K, Pearse MJ, d'Apice AJ: **Targeting gene expression to endothelium in transgenic animals: a comparison of the human ICAM-2, PECAM-1 and endoglin promoters.** *Xenotransplantation* 2003, **10**:223-231.
63. Korn T, Muller R, Kontermann RE: **Bispecific single-chain diabody-mediated killing of endoglin-positive endothelial cells by cytotoxic T lymphocytes.** *J Immunother* 2004, **27**:99-106.
64. Savontaus MJ, Sauter BV, Huang TG, Woo SL: **Transcriptional targeting of conditionally replicating adenovirus to dividing endothelial cells.** *Gene Ther* 2002, **9**:972-979.
65. Bredow S, Lewin M, Hofmann B, Marecos E, Weissleder R: **Imaging of tumour neovasculature by targeting the TGF-beta binding receptor endoglin.** *Eur J Cancer* 2000, **36**:675-681.
66. Costello B, Li C, Duff S, Butterworth D, Khan A, Perkins M, Owens S, Al-Mowallad AF, O'Dwyer S, Kumar S: **Perfusion of 99Tcm-labeled CD105 Mab into kidneys from patients with renal carcinoma suggests that CD105 is a promising vascular target.** *Int J Cancer* 2004, **109**:436-441.

67. Seon BK, Matsuno F, Haruta Y, Kondo M, Barcos M: **Long-lasting complete inhibition of human solid tumors in SCID mice by targeting endothelial cells of tumor vasculature with anti-human endoglin immunotoxin.** *Clin Cancer Res* 1997, **3**:1031-1044.
68. Matsuno F, Haruta Y, Kondo M, Tsai H, Barcos M, Seon BK: **Induction of lasting complete regression of preformed distinct solid tumors by targeting the tumor vasculature using two new anti-endoglin monoclonal antibodies.** *Clin Cancer Res* 1999, **5**:371-382.
69. Tabata M, Kondo M, Haruta Y, Seon BK: **Antiangiogenic radioimmunotherapy of human solid tumors in SCID mice using ¹²⁵I-labeled anti-endoglin monoclonal antibodies.** *Int J Cancer* 1999, **82**:737-742.
70. Takahashi N, Haba A, Matsuno F, Seon BK: **Antiangiogenic therapy of established tumors in human skin/severe combined immunodeficiency mouse chimeras by anti-endoglin [CD105] monoclonal antibodies, and synergy between anti-endoglin antibody and cyclophosphamide.** *Cancer Res* 2001, **61**:7846-7854.
71. Griffioen AW, Damen CA, Blijham GH, Groenewegen G: **Endoglin/CD105 may not be an optimal tumor endothelial treatment target.** *Breast Cancer Res Treat* 1996, **39**:239-242.
72. Balza E, Castellani P, Zijlstra A, Neri D, Zardi L, Siri A: **Lack of specificity of endoglin expression for tumor blood vessels.** *Int J Cancer* 2001, **94**:579-585.
73. Seon BK: **Expression of endoglin [CD105] in tumor blood vessels.** *Int J Cancer* 2002, **99**:310-311.
74. Grisanti S, Canbek S, Kaiserling E, Adam A, Lafaut B, Gelisken F, Szurman P, Henke-Fahle S, Oficjalska-Mlynaczak J, Bartz-Schmidt KU: **Expression of endoglin in choroidal neovascularization.** *Exp Eye Res* 2004, **78**:207-213.
75. Kumar S, Ghellal A, Li C, Byrne G, Haboubi N, Wang JM, Bundred N: **Breast carcinoma: vascular density determined using CD105 antibody correlates with tumor prognosis.** *Cancer Res* 1999, **59**:856-861.
76. Dales JP, Garcia S, Bonnier P, Duffaud F, Andrac-Meyer L, Ramuz O, Lavaut MN, Allasia C, Charpin C: **CD105 expression is a marker of high metastatic risk and poor outcome in breast carcinomas. Correlations between immunohistochemical analysis and long-term follow-up in a series of 929 patients.** *Am J Clin Pathol* 2003, **119**:374-380.
77. Brewer CA, Setterdahl JJ, Li MJ, Johnston JM, Mann JL, McAsey ME: **Endoglin expression as a measure of microvessel density in cervical cancer.** *Obstet Gynecol* 2000, **96**:224-228.
78. Li C, Gardy R, Seon BK, Duff SE, Abdalla S, Renehan A, O'Dwyer ST, Haboubi N, Kumar S: **Both high intratumoral microvessel density determined using CD105 antibody and elevated plasma levels of CD105 in colorectal cancer patients correlate with poor prognosis.** *Br J Cancer* 2003, **88**:1424-1431.
79. Saad RS, Liu YL, Nathan G, Celebrezze J, Medich D, Silverman JF: **Endoglin [CD105] and vascular endothelial growth factor as prognostic markers in colorectal cancer.** *Mod Pathol* 2004, **17**:197-203.
80. Salvesen HB, Gulluoglu MG, Stefansson I, Akslen LA: **Significance of CD105 expression for tumour angiogenesis and prognosis in endometrial carcinomas.** *APMIS* 2003, **111**:1011-1018.
81. Saad RS, Jasnosz KM, Tung MY, Silverman JF: **Endoglin [CD105] expression in endometrial carcinoma.** *Int J Gynecol Pathol* 2003, **22**:248-253.
82. Straume O, Akslen LA: **Expression of vascular endothelial growth factor, its receptors [FLT-1, KDR] and TSP-1 related to microvessel density and patient outcome in vertical growth phase melanomas.** *Am J Pathol* 2001, **159**:223-235.
83. Adam M, Schmidt D, Wardemann E, Wernert N, Albers P: **Angiogenetic protooncogene ets-1 induced neovascularization is involved in the metastatic process of testicular germ cell tumors.** *Eur Urol* 2003, **44**:329-336.
84. Tanaka F, Otake Y, Yanagihara K, Kawano Y, Miyahara R, Li M, Yamada T, Hanaoka N, Inui K, Wada H: **Evaluation of angiogenesis in non-small cell lung cancer: comparison between anti-CD34 antibody and anti-CD105 antibody.** *Clin Cancer Res* 2001, **7**:3410-3415.
85. Yagasaki H, Kawata N, Takimoto Y, Nemoto N: **Histopathological analysis of angiogenic factors in renal cell carcinoma.** *Int J Urol* 2003, **10**:220-227.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

