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Dysregulation of *miR-200s* clusters as potential prognostic biomarkers in acute myeloid leukemia

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Abstract

Background: Increasing studies showed that *miR-200* family (*miR-200s*) clusters are aberrantly expressed in multiple human cancers, and *miR-200s* clusters function as tumor suppressor genes by affecting cell proliferation, self-renewal, differentiation, division and apoptosis. Herein, we aimed to investigate the expression and clinical implication of *miR-200s* clusters in acute myeloid leukemia (AML).

Methods: RT-qPCR was performed to detect expression of *miR-200s* clusters in 19 healthy donors, 98 newly diagnosed AML patients, and 35 AML patients achieved complete remission (CR).

Results: Expression of *miR-200a/200b/429* cluster but not *miR-200c/141* cluster was decreased in newly diagnosed AML patients as compared to healthy donors and AML patients achieved CR. Although no significant differences were observed between *miR-200s* clusters and most of the features, low expression of *miR-200s* clusters seems to be associated with higher white blood cells especially for *miR-200a/200b*. Of the five members of *miR-200s* clusters, low expression of *miR-200b/429/200c* was found to be associated with lower CR rate. Logistic regression analysis further revealed that low expression of *miR-200b/429/200c* was associated with shorter OS, whereas *miR-200a/141* had a trend. Moreover, multivariate analysis of Cox regression models confirmed the independently prognostic value of *miR-200b* expression for OS in AML.

Conclusions: Expression of *miR-200a/200b/429* cluster was frequently down-regulated in AML, and low expression of *miR-429* as an independent risk factor for CR, whereas low expression of *miR-200b* as an independent prognostic biomarker for OS.

Keywords: miR-200, Expression, Prognosis, Acute myeloid leukemia

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Background

Acute myeloid leukemia (AML) is a highly heterogeneous malignant hematological disorder with complex molecular pathophysiology. Although the treatment strategies against AML have been updated in the past decades, the majority of patients eventually succumb to relapse after induction chemotherapy [1]. Clinical outcome of AML remains unsatisfactory especially in those with specific karyotypes/biomarkers such as inv(3)(g21g26.2), t(6;9) (p23; q34), 11q abnormalities other than t(9;11), -5/ del(5q), -7, TP53 mutations, FLT3-ITD mutations, C-KIT mutations, WT1 overexpression, and BAALC overexpression [2–4]. The development of effective therapeutic options against AML relies on mechanistic understanding of AML biology, especially in molecular regulators of AML pathogenesis and molecular predictor of AML prognosis [5].

MicroRNAs, a class of small (19–22 nucleotides) single-stranded RNAs, negatively regulate various genes by targeting 3'-untranslated region (3'-UTR) of mRNAs, thereby facilitating translational silencing or degradation of targeted genes [6]. Mounting evidences have implicated that microRNAs play crucial roles in regulating many fundamental and biological processes including cancer development [7]. Moreover, microRNAs have been reported as novel biomarkers for diagnosis and prognosis, and regarded as potential therapeutic targets in AML [8]. For instance, recent studies implicated that several microRNAs such as *miR-216b, miR-362-5p, miR-217*, and *miR-193b* were prognosis-related predictors in AML and may involve in AML biology [9–12].

The *miR-200* family (*miR-200s*) clusters includes five members (*miR-200a*, *miR-200b*, *miR-200c*, *miR-141*, and *miR-429*) and can be divided into two clusters (*miR-200a/b/429* cluster and *miR-200c/141* cluster) based on chromosomal location (chromosome 1p36 and chromosome 12p13) [13]. Numerous studies showed that *miR-200s* clusters are aberrantly expressed in multiple human cancers, and *miR-200s* clusters function as tumor suppressor genes by affecting cell proliferation, self-renewal, differentiation, division and apoptosis [14]. Although the tumor-suppressive roles of *miR-200s* clusters have also been reported in solid tumors with prognostic value [14, 15], the expression and clinical implication of *miR-200s* clusters in AML remains poorly revealed.

In this study, we investigated expression of *miR-200s* clusters in AML patients except for acute promyelocytic leukemia (APL), and found that low expression of *miR-200s* clusters acted as potential prognostic biomarkers in AML.

Methods

Patients and treatment

A total of 98 de novo AML patients except for APL and 19 healthy donors were enrolled in this study. Bone marrow (BM) was collected from all the patients at diagnosis time as well as 35 patients at complete remission (CR) time. AML was diagnosed based on the French-American-British (FAB) and 2016 revised World Health Organization (WHO) criteria [16, 17]. All the patients received chemotherapy as reported [18]. Induction chemotherapy therapy was 1-2 courses of daunorubicin combined with cytarabine. Subsequent consolidation treatment after CR for younger patients included highdose cytarabine, mitoxantrone with cytarabine, and homoharringtonine combined with cytarabine, whereas for older patients received in an individualized manner decided by the physicians, such as CHG protocol (cytarabine, homoharringtonine, and G-CSF). This study was approved by the Ethics Committee of the Affiliated People's Hospital of Jiangsu University, and written informed consents were informed and signed by all participants in accordance with the Declaration of Helsinki Principles.

Cytogenetic analysis and mutation detection

BM cells were harvested after 1–3 days of unstimulated culture in RPMI 1640 medium (BOSTER, Wuhan, China) containing 20% fetal calf serum (ExCell Bio, Shanghai, China). Cytogenetics for AML patients were analyzed at the newly diagnosis time by conventional R-banding method and karyotype risk was classified according to reported previously [19, 20]. Hotspot mutations in *NPM1, C-KIT, DNMT3A, N/K-RAS, IDH1/2, U2AF1, SRSF2* and *SETBP1* were detected by high-resolution melting analysis [21–25], whereas mutations in *FLT3*-ITD and *CEBPA* were examined by DNA sequencing [26].

RNA isolation and reverse transcription

BM mononuclear cells (BMMNCs) were extracted as reported using Lymphocyte Separation Medium (Absin, Shanghai, China) [27]. According to the manufacturer's protocols, RNA was extracted from BMMNCs using the mirVana miRNA isolation kit (Ambion, Austin, TX, USA), and was synthesized to cDNA by reverse transcription using MiScript Reverse Transcription Kit (Qiagen, Duesseldorf, Germany).

Real-time quantitative PCR

The level of *miR-200s* clusters was detected by real-time quantitative PCR (RT-qPCR) using miScript SYBR green PCR kit (Qiagen, Duesseldorf, Germany). The primers were *miR-200s* specific (Additional file 1: Table S1) and the manufacturer-provided miScript universal primer

(Qiagen, Duesseldorf, Germany). The programs for RTqPCR reactions were performed as reported [28]. *U6* small nuclear RNA was selected as the endogenous normalizer detected by RT-qPCR using 2× SYBR Green PCR Mix (Multisciences, Hangzhou, China). Relative *miR-200s* level was calculated by $2^{-\Delta\Delta CT}$ method. The healthy donors that possessed the minimal ΔCT between *miR-200s* (each member) and *U6* expression was selected as control, and was defined as 100% expression.

Statistical analysis

Mann–Whitney's U test was carried to compare the difference of continuous variables between two groups, whereas Pearson Chi square analysis/Fisher exact test were applied to compare the difference of categorical variables between two groups. The impact of *miR*-200s clusters expression on overall survival (OS) was analyzed by Kaplan–Meier analysis, and Cox regression models (univariate and multivariate analyses) were further used to determine the independently prognostic value of *miR*-200s clusters expression on CR was determined by Logistic regression analysis (univariate and multivariate and multivariate and multivariate and multivariate and multivariate and multivariate and set of *miR*-200s clusters expression on CR was determined by Logistic regression analysis (univariate and multivariate analyses). All tests were two sided, and P < 0.05 was

defined as statistically significant. SPSS software 20.0 and GraphPad Prism 5.0 was used to conduct the statistical analyses in this study.

Results

Expression of miR-200s in AML

We analyzed *miR-200s* clusters expression in BM from 19 healthy donors, 98 AML patients, and 35 AML patients achieved CR by RT-qPCR. As presented in Fig. 1, expression of *miR-200a/200b/429* clusters but not *miR-200c/141* clusters was significantly decreased in AML patients as compared to healthy donors and AML patients achieved CR.

Relationship between miR-200s and clinical features in AML

To investigate clinical implication of *miR-200s* clusters expression, the whole-cohort patients were classified into two groups (high and low *miR-200s* clusters expression) based on the median level of each member of *miR-200s* clusters, respectively. We analyzed the association between each member of *miR-200s* clusters expression and clinic-pathologic features including gender, age,



Fig. 1 Expression of *miR-200s* clusters in controls, newly diagnosed AML patients and AML patients achieved CR. **a** For *miR-200a*; **b** For *miR-200b*; **c** For *miR-429*; **d** For *miR-200s*; **e** For *miR-141*. The distributions of the *miR-200s* clusters expression in controls, newly diagnosed AML patients and AML patients achieved CR were presented with scatter plots. The median level of *miR-200s* clusters expression in each group was shown with horizontal line

features in AML patients	
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Correlation of <i>miR-200</i> s cl	
Table 1(

Patient's	<i>miR-200а</i> ехр	ression		miR-200b expi	ression		miR-429 expre	ession		<i>miR-200</i> c exp	ression		miR-141 expr	ession	
features	Low (n = 49)	High (n = 49)	ط	Low (n = 49)	High (n = 49)	ط	Low (n = 49)	High (n = 49)	٩	Low (n= 49)	High (n = 49)	ط	Low (n= 49)	High (n=49)	٩
Sex (male/ female)	36/13	23/26	0.013	32/17	27/22	0.409	31/18	28/21	0.680	32/17	27/22	0.409	31/18	28/21	0.680
Age (years)	58 (21–81)	61 (18–87)	0.842	60 (18-81)	59 (18–87)	0.831	60 (21–81)	59 (18–87)	0.741	60 (18–81)	59 (18–87)	0.743	60 (18–81)	59 (18–87)	1.000
WBC (× 10 ⁹ /L)	38.7 (1.3–528.0)	9.6 (1.1–130.2)	0.001	35.5 (1.3–528.0)	9.6 (1.1–130.2)	0.041	34.5 (1.3–528.0)	13.3 (1.1–130.2)	660.0	34.9 (1.3–528.0)	13.2 (1.1–130.2)	0.094	35.5 (1.3–528.0)	10.2 (1.1–116.6)	060.0
Hemoglobin (g/L)	78 (53–138)	76.5 (32–144)	0.085	80 (53–138)	76.5 (32–144)	0.330	77 (32–138)	78 (34–134)	0.940	84 (53–138)	76 (32–144)	0.124	88 (53–138)	74.5 (32–144)	0.024
Platelets $(\times 10^{9}/L)$	37 (3–447)	47 (4–264)	0.558	37 (3-447)	46 (4–264)	0.460	36 (3–447)	47 (4–264)	0.512	30 (5-447)	49 (3–264)	0.233	40 (5–125)	46 (3-447)	0.769
BM blasts (%)	60% (20–99%)	58% (20–95%)	0.892	63% (20–99%)	58% (21–95%)	0.651	60% (20–98%)	59% (20–99%)	0.757	63% (20–99%)	54% (20–95%)	0.226	62% (20–98%)	5 <i>7</i> % (20–99%)	0.598
FAB sub- types			0.660			0.945			0.681			0.827			006.0
MO	-	0		-	0		0	-		1	0		<i>—</i>	0	
M1	4	2		Ω	3		C	3		2	4		2	4	
M2	26	24		23	27		22	28		23	27		24	26	
M4	12	15		14	13		15	12		15	12		15	12	
M5	9	9		7	5		~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	4		7	5		9	9	
M6	0	2		-	1		_	<i>—</i>		1	—		1	-	
Karyotypes			0.982			0.813			0.606			0.948			0.578
Normal	25	28		23	30		23	30		24	29		24	29	
t(8;21)	5	5		5	5		9	4		5	5		9	4	
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t(9;22)	-	0		,	0		0	, –		-	0		-	0	
Complex	00	7		6	9		10	5		00	7		7	80	
Others	9	5		9	5		9	5		9	5		5	9	
No data	0	1		0	-		0			0	-		0	-	
Gene mutatio	Sh														
CEBPA (土)	7/33	6/38	0.765	5/38	8/33	0.376	8/36	5/35	0.555	7/34	6/37	0.768	6/37	7/34	0.768
NPM1 (土)	5/35	4/40	0.730	3/40	6/35	0.307	4/40	5/35	0.730	5/36	4/39	0.735	4/39	5/36	0.735
<i>FLT3-</i> ITD (±)	6/34	3/41	0.298	6/37	3/38	0.484	6/38	3/37	0.488	5/36	4/39	0.735	6/37	3/38	0.484

Table 1 (co	intinued)														
Patient's	miR-200a €	sxpression		<i>miR-200b</i> e	xpression		miR-429 ex	pression		<i>miR-200c</i> e)	cpression		miR-141 exp	oression	
features	Low (n = 49)	High (n = 49)	٩	Low (n = 49)	High (n = 49)	٩	Low (n = 49)	High (n = 49)	ط	Low (n=49)	High (n = 49)	٩	Low (n=49)	High (n=49)	ط
C-KIT (土)	1/39	1/43	1.000	1/42	1/40	1.000	1/43	1/39	1.000	1/40	1/42	1.000	1/42	1/40	1.000
N/K-RAS (±)	4/36	6/38	0.741	3/40	7/34	0.190	5/39	5/35	1.000	6/35	4/39	0.515	6/37	4/37	0.739
(∓) <i>7/1H0</i> I	1/39	3/41	0.618	3/40	1/40	0.616	4/40	0/40	0.118	4/37	0/43	0.052	2/41	2/39	1.000
DNMT3A (土)	4/36	2/42	0.418	4/39	2/39	0.676	4/40	2/38	0.678	3/38	3/40	1.000	3/40	3/38	1.000
U2AF1 (土)	2/38	2/42	1.000	1/42	3/38	0.354	1/43	3/37	0.343	2/39	2/41	1.000	3/40	1/40	0.616
SRSF2 (土)	1/39	3/41	0.618	1/42	3/38	0.354	1/43	3/37	0.343	1/40	3/40	0.616	1/42	3/38	0.354
SETBP1 (±)	2/38	0/44	0.224	2/41	0/41	0.494	2/42	0/40	0.495	2/39	0/43	0.235	2/41	0/41	0.494
CR (±)	16/33	23/26	0.215	14/35	25/24	0.038	14/35	25/24	0.038	14/35	25/24	0.038	15/34	24/25	0.098
WBC white bloc	od cells, <i>BM</i> bo	ne marrow, FAB	French–Ar	merican–British	ו classification, C	R complet	e remission								

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Variables	Complete remission				Overall survival			
	Univariate analysis		Multivariate analysis	5	Univariate analysis		Multivariate analysis	;
	OR (95% CI)	Р	OR (95% CI)	Р	HR (95% CI)	Р	HR (95% CI)	Р
miR-200a	0.548 (0.242–1.244)	0.150	1.029 (0.285–3.722)	0.965	0.662 (0.415-1.022)	0.082	1.425 (0.651–3.120)	0.376
miR-200b	0.384 (0.167–0.885)	0.025	0.823 (0.199–3.401)	0.788	0.511 (0.319–0.819)	0.005	0.524 (0.305–0.902)	0.020
miR-429	0.384 (0.167–0.885)	0.025	0.331 (0.128–0.858)	0.023	0.558 (0.350–0.891)	0.015	0.820 (0.325–2.073)	0.675
miR-200c	0.384 (0.167–0.885)	0.025	0.977 (0.149–6.400)	0.981	0.606 (0.380–0.965)	0.035	0.649 (0.190–2.217)	0.491
miR-141	0.460 (0.201-1.050)	0.065	0.594 (0.192–1.833)	0.364	0.695 (0.437–1.104)	0.123	1.152 (0.582–2.279)	0.684
Age	4.229 (1.742–10.266)	0.001	4.555 (1.715–12.095)	0.002	2.046 (1.282–3.266)	0.003	1.732 (1.033–2.902)	0.037
WBC	2.367 (1.015–5.520)	0.046	1.846 (0.715–4.767)	0.206	2.002 (1.253–3.199)	0.004	1.560 (0.925–2.629)	0.095
Karyotype	3.108 (1.338–7.220)	0.008	2.862 (1.164–7.042)	0.022	1.875 (1.295–2.715)	0.001	1.874 (1.210–2.902)	0.005
CEBPA mutations	0.526 (0.160–1.731)	0.290			0.870 (0.413–1.829)	0.713		
NPM1 mutations	0.833 (0.207–3.358)	0.798			1.200 (0.516–2.793)	0.672		
FLT3-ITD mutations	0.833 (0.207–3.358)	0.798			0.935 (0.403–2.170)	0.876		
C-KIT mutations	0.673 (0.041–11.150)	0.783			0.479 (0.066–3.458)	0.465		
N/K-RAS mutations	3.048 (0.605–15.343)	0.177			1.311 (0.621–2.770)	0.478		
IDH1/2 mutations	Undetermined	0.999			4.671 (1.637–13.326)	0.004	6.662 (1.757–25.268)	0.005
DNMT3A mutations	1.391 (0.240–8.057)	0.712			1.590 (0.634–3.987)	0.323		
U2AF1 mutations	Undetermined	0.999			2.791 (0.987–7.890)	0.053	5.130 (1.714–15.355)	0.003
SRSF2 mutations	Undetermined	0.999			1.934 (0.693–5.400)	0.208		
SETBP1 mutations	0.673 (0.041–11.150)	0.783			0.637 (0.088–4.613)	0.656		

Table 2 Univariate and multivariate analyses of variables for overall survival in AML patients

OR odd ratio, *HR* hazard ratio, *Cl* confidence interval. Variables including *miR-200s* cluster expression (Low vs. High), age (≤ 60 vs. > 60 years), WBC ($\geq 30 \times 10^9$ vs. < 30×10^9 /L), karyotype (favorable vs. intermediate vs. poor), and gene mutations (mutant vs. wild-type). Multivariate analysis includes variables with *P* < 0.200 in univariate analysis

white blood cell (WBC) counts, hemoglobin content, platelet counts, blasts (%), FAB subtypes, karyotypes, and common gene mutations. As shown in Table 1, no significant differences were observed between *miR-200s* clusters expression and most of the features. However, low expression of *miR-200s* clusters seems to be associated with higher WBC counts especially for *miR-200a*/200b (P=0.001 and 0.041, respectively). In addition, low expression of *miR-200a* was related to male, whereas low expression of *miR-141* was correlated with higher hemoglobin content (P=0.013 and 0.024, respectively).

Prognostic value of miR-200s in AML

To observe the impact of *miR-200s* clusters expression on clinical outcome in AML, we first determined the association of each member of *miR-200s* clusters expression with CR. Of the five members of *miR-200s* clusters, low expression of *miR-200b/429/200c* was found to be associated with lower CR rate (Table 1, all P=0.038). Additionally, Logistic regression analysis was further performed to confirm and verify the effect of *miR-200s* clusters' expression on CR, and revealed low expression of *miR-429* as an independent risk factor for CR in AML (Table 2, P=0.023). We next evaluated the correlation of each member of *miR-200s* clusters expression with survival. Based on Kaplan–Meier analysis, low expression of *miR-200b/429/200c* was associated with shorter OS, whereas *miR-200a/141* had a trend (Fig. 2). In addition, we also analyzed the impact of composite members of *miR-200s* clusters expression on OS by Kaplan–Meier analysis as shown in Fig. 3.

Since miR-200s clusters expression was associated with well-established prognostic factor such as WBC counts, we further conducted a Cox regression model adjusting for prognosis-related factors (age, WBC counts, karyo-typic classifications, and gene mutations) for OS. Results showed that low expression of miR-200b acted as an independent prognostic biomarker for OS (P=0.020, Table 2).

Discussion

In the current study, we for the first time investigated expression of *miR-200s* clusters in AML, and revealed that most of the members of *miR-200s* clusters were down-regulated in de novo AML patients. Recently, Li et al. revealed that introduction of a pre-*miR-200c* reduced the expression of ZEB2 protein and inhibited the proliferation of human leukemia cell lines (HL-60,



MOLM-13, and THP-1), and mouse miR-200c significantly impaired the proliferation of mouse leukemia cells [29]. Taken together, these results emphasized the crucial role of miR-200s clusters in leukemogenesis. Although the biological role of miR-200s clusters in AML was less studied, tumor suppressor roles of miR-200s clusters have been identified in a variety of human solid cancers, such as bladder cancer, gastric cancer, colorectal cancer, breast cancer, ovarian cancer, endometrial cancer, pancreatic cancer, gliomas, hepatocellular carcinoma, and lung cancer [14, 30]. The *miR-200s* clusters were reported as key inhibitors of epithelial-to-mesenchymal transition by directly targeting transcriptional repressors of E-cadherin, ZEB1, and ZEB2 [13]. Moreover, miR-200s clusters also played crucial roles in the repression of cancer stem cells self-renewal and differentiation, modulation of cell division and apoptosis, and reversal of chemoresistance [14, 30]. Notably, in some other hematological malignancies, expression or biological role of miR-200s clusters has been preliminary studied. For instance, Choi et al. reported that miR-200c was decreased in patients with myelodysplastic syndrome (MDS) [31]. González-Gugel

et al. revealed that down-regulation of mmu-*miR-30a* and mmu-*miR-141* as well as hsa-*miR-193b* clearly contributed to enhance the expression of *Smoothened* (*SMO*) gene in mouse and human lymphomas and, subsequently, to activate the GLI/Hh signalling [32].

In addition to basic research before, it has been noted that low expression of miR-200s clusters could correlate with adverse clinical outcome and serve as a prognostic biomarker for various cancer patients [15]. Although the potential prognostic value of miR-200s clusters in several human cancers remains controversial, a recent meta-analysis demonstrated that lower tissue expression of miR-200s clusters' members were associated with poor OS and progression-free survival, whereas lower expression of circulating miR-200s clusters' members were correlated with favorable prognosis [15]. From our study, we showed the negative effect of low expression of miR-200s clusters on AML chemotherapy response and survival. Moreover, multivariate analysis showed that low expression of miR-429 as an independent risk factor for CR, whereas low expression of miR-200b as an independent prognostic biomarker for OS in AML. Due to some



two members of *miR-200s* clusters abnormalities). **E** OS analyzed between two groups (equal or more than two members of *miR-200s* clusters abnormalities). **F** OS analyzed between two groups (equal or more than one member of *miR-200s* clusters abnormalities). **F** OS analyzed between two groups (equal or more than one member of *miR-200s* clusters abnormalities). **F** OS analyzed between two groups (equal or more than one member of *miR-200s* clusters abnormalities). **F** OS analyzed between two groups (equal or more than one member of *miR-200s* clusters abnormalities).

limitations in this study (such as patients numbers, treatment regimens, and single center), prospective studies are needed to verify our results before *miR-200s* clusters expression could be used routinely as a promising biomarker for risk stratification in AML.

Conclusion

Expression of *miR-200a/200b/429* cluster was frequently down-regulated in AML, and low expression of *miR-429* as an independent risk factor for CR, whereas low expression of *miR-200b* as an independent prognostic biomarker for OS.

Additional file

Additional file 1: Table S1. The primer sequences for miR-200s clusters.

Abbreviations

AML: acute myeloid leukemia; 3'-UTR: 3'-untranslated region; APL: acute promyelocytic leukemia; BM: bone marrow; CR: complete remission; BMMNCs: BM mononuclear cells; FAB: French–American–British; WHO: World Health Organization; RT-qPCR: real-time quantitative PCR; OS: overall survival; WBC: white blood cell.

Authors' contributions

JQ and JL conceived and designed the experiments; JZ and LZ performed the experiments; JZ and TZ analyzed the data; YG, WZ and DW collected the clinical data; JM, XW and HG offered technique support; JZ wrote the paper. All authors read and approved the final manuscript.

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None.

Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Consent for publication

Written informed consents were obtained from all enrolled individuals prior to their participation.

Ethical approval and consent to participate

The present study approved by the Ethics Committee and Institutional Review Board of the Affiliated People's Hospital of Jiangsu University.

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